

# Multidisciplinary Advances in Veterinary Science

ISSN: 2573-3435

**Research Article** 

# Hematological and Serological Changes in Neonatal Diarrheic Calves Infected with Bovine Rotavirus

# Shama Ranjan Barua\*, Tofazzal Md Rakib, Shubhagata Das, Md Masuduzzaman, Mohammad Alamgir Hossain and Sharmin Chowdhury

Department of Pathology and Parasitology, Chittagong Veterinary and Animal Sciences University, Khulshi, Chittagong 4225, Bangladesh

\*Corresponding Author: Shama Ranjan Barua, Department of Pathology and Parasitology, Chittagong Veterinary and Animal Sciences University, Khulshi, Chittagong 4225, Bangladesh.

Received: May 30, 2018; Published: June 05, 2018

# Abstract

Bovine rotavirus A is a leading cause of diarrhea in calves. Diarrhea causes excessive water and electrolyte loss through the gastrointestinal (GI) tract and subsequently produces dehydration, and in acute cases can lead to death. The aim of the study was to evaluate haemato-biochemical changes occurred in diarrheic calves as well as rotavirus affected diarrheic calves. The study was undertaken in calves between 1-45 days old at Chittagong metropolitan area in Chittagong, Bangladesh during the period from January 2017 to June 2017. Fifty blood samples from 40 diarrheic and 10 non-diarrheic cases were collected and subjected to analysis of hematological and serological parameters. Hematological values in both rotavirus positive and negative cases had no significant differences except increased number of lymphocytes (75.58%; SD: 7.34; 95% CI: 71.81-79.36) which clearly indicate viral causation. Serological profile revealed increase in chloride level (115.05%; SD: 19.18; 95% CI: 108.83-121.27) in rotavirus positive diarrheic calves as compared to rotavirus negative cases and suggested the presence of hyperchloremia in diarrheic calves.

Keywords: Hematology; Serology; Rotavirus Diarrhea; Calves

Volume 2 Issue 3 June 2018

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## Introduction

Neonatal calf diarrhea is a major health problem in dairy hard management. It has a multifactorial etiology including viruses, bacteria, protozoa and also management factors [1,2]. In respect to the enter pathogens, Escherichia coli K99 (*E. coli*), Coronavirus, Cryptosporidium parvum (C. parvum) and Rotavirus causes 75–95% of intestinal infections in young calves and especially rotavirus accounts for 27–36% [3-5]. Rotavirus is responsible for impairment of the mature enterocytes of the villi (primarily in the middle and terminal segment) of the small intestine and enhances its exfoliation [6]. As a result disturbed water and electrolyte absorption is observed, which, in turn, causes its accumulation in the gut lumen resulting diarrhea [6]. It leads to morbidity and mortality, delayed growth rate, higher age at first calving that consequence to direct and indirect economic losses in dairy farming [1, 7-9].

Rapid diagnosis of etiology and estimation of physiological alteration are important factors for proper treatment and control of diseases. Hematological and biochemical variables are most frequently used as diagnostic tool for taking medical decision. Analyses of blood parameters are very helpful to have an insight in metabolic and health status of animal. It is also very useful to compare the values obtained from ill animal with normal values in healthy animals during diagnostic procedure [10]. The changes in biochemical and hematological constituents are important indicators of the physiological or pathological state of the animal. Blood examination is also performed for screening of general health [11]. Diarrheic animals loose fluid, becomes rapidly dehydrated and are suffered from electrolyte loss, acidosis. Infectious agents may cause initial damage to the intestine but death from scours usually results from dehydration, acidosis and loss of electrolytes [12].

Packed cell volume (PCV) may increase in diarrheic calves that indicates fluid loss from vascular compartment [13], and oral rehydration solution therapy along with other therapeutic measure were effective in bringing back the hydration status of diarrheic calves to normal [14]. Therefore, packed cell volume (PCV) could be an important indicator of change in extra cellular fluid (ECF) volume during diarrhea. Dehydration in calves that have diarrhea is accompanied by large decreases in the extracellular fluid volume along with small increases in intracellular fluid volume [15,16]. In general, significant increase in hematological parameters as Hemoglobin (Hb), packed cell volume (PCV) and total erythrocyte counts (TEC) due to haemoconcentration are related with dehydration [17]. Moreover, some biochemical alterations such as hypoglycemia [17], Hyperproteinaemia and hyperalbuminaemia [18], hyperkalemia, hyperchloremia and hypernatremia[19] were also found associated with diarrhea.

The study was designed to estimate the hematological and serological values in diarrheal/non-diarrheal, rotavirus positive/negative cases in calves that might support the diagnosis of the suspected etiology.

#### Methodology

#### Study area and sample size

The study was undertaken on cattle calves under the age of 45 days exhibiting symptoms of diarrhea in selected dairy farms in Chittagong. A total of 50 samples (blood and faeces), among which 40 from the calves showing diarrhea and 10 samples without diarrhea were collected.

#### Sample collection

The fecal samples were collected directly from rectum using latex gloves. In addition, five to ten ml of blood from each of the calves was collected from the jugular vein in aseptic conditions using labeled sterile disposable syringes. About half of the blood was transferred immediately after collection to the sterilized vial containing ethylene di-amine tetra acetic acid (EDTA) solution and half of the blood was transferred to sterilized labeled test tubes for serum. All the samples were transported using cool box to the Clinical Pathology laboratory, Department of Pathology and Parasitology, Chittagong Veterinary and Animal Sciences University, Khulshi, Chittagong.

#### Sample processing

A 10% fecal suspension of individual sample was prepared in phosphate buffered saline (P<sup>H</sup> 7.2) and clarified by centrifugation at 13,000 rpm at 4°C for 10 minutes. The supernatant was separated and stored at -70°C until RNA extraction for real time RT-PCR. After coagulation, coagulated blood was centrifuged in 3000 rpm for 15 minutes. Serum was transferred to the eppendorf tube by using micropipette. The obtained serum samples were stored in -20°C at laboratory for biochemical test.

#### Laboratory analysis of blood and serum samples

Hematological analysis of total erythrocyte count (TEC/RBC), total leukocyte count (TLC/WBC), differential leukocyte count (DLC), packed cell volume (PCV), hemoglobin concentration (Hb) were done as per standard procedures described by Nemi [20] and Sharma and Singh [21]. Consequently, serological investigation was conducted following proper aseptic measures at the time of serum analysis

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357

358

in the laboratory. Serum was placed in normal temperature approximately 30 minutes before the analysis. Serum samples were evaluated for electrolyte parameters by an automated biochemical analyzer (HumalizerR-3000).

#### Detection of rotavirus by real time RT- PCR

All the fecal samples were used for viral RNA extraction. Rotavirus RNA was extracted from the fecal suspension using QIAamp Viral RNA mini kit (Qiagen/Westburg, Leusden, Netherlands) following the manufacturer's instructions. The extracted RNA were stored at -80°C. Subsequently, the real time RT-PCR reaction was performed by using a QIAGEN One Step RT-PCR Kit (QIAGEN) for the confirmation of bovine rotavirus A. The real time RT-PCR assay was performed by using an ABI 7500 thermo cycler (Applied Biosystems). The TaqMan® assay primers and probe sequences and thermal cycles were selected for testing according to previously described by Jothikumar, Kang [22].

#### Statistical analysis

The laboratory data was stored in Microsoft Excell-2007 and Statistical differences were calculated according to the Student t-test with significance level at P < 0.05 using the procedure of (SAS, 2004). Descriptive statistics analysis was done to measure the mean, SD, 95% confidence interval (CI) and P value of different parameters. The two sample paired t-test was done to compare the diarrhea/non-diarrhea, rotavirus positive/negative and diarrhea with rotavirus positive/negative variation.

## **Results**

The blood samples were subjected to estimation of hematological parameters including Hemoglobin, packed cell volume, total erythrocyte count, total leukocyte count and differential leukocyte count and serological parameters including sodium, potassium and chloride. All the parameter estimates has been compiled in Table 3-8 and compared with the reference values of Table 1&2.

Parameters	Normal Range (Calves)	Diarrheic Calves
Haemoglobin (g/dl)	11.3 (9.5-13.5)	$12.2\pm0.221$
PCV (%)	37.5 (36.0-49.0)	$44.2\pm0.692$
TEC (million/cm <sup>3</sup> )	8.1	$8.54\pm0.316$
TLC (thousands/cm <sup>3</sup> )	9.7	$12.4\pm0.295$
Neutrophils (%)	37 (12-38)	$46.8\pm0.748$
Eosinophils (%)	1.9 (2-30)	$1.7\pm0.482$
Basophils (%)	0.6 (0-2)	$0.5\pm0.147$
Lymphocytes (%)	68.6 (33-87)	$51.6\pm0.514$
Monocytes (%)	3.4 (1-5)	$2.72\pm0.687$

Courtesy by Mailk, Kumar (36)

Table 1: Hematological profiles of normal and diarrheic calves.

Parameters	Normal range (Calves)#	Diarrheic calves##
Na (mEq/litre)	$144\pm2.9$	$126.16 \pm 1.815$
K (mEq/litre)	$5.95\pm0.48$	$5.6\pm0.084$
Cl- (mEq/litre)	$102.3\pm2.7$	$122.47 \pm 1.943$

# Klinkon and Ježek (10); ## Mailk, Kumar (36)

Table 2: Biochemical profile of normal and diarrheic calves.

Variable	Infection status	Mean	SD	95% CI	P-value (Ttest)
Hemoglobin (g/dl)	Non-diarrhoea	8.01	2.01	6.66-9.36	
	Diarrhoea	10.17	2.30	9.43-10.92	3.05
PCV (%)	Non-diarrhoea	31.45	9.71	24.93-37.98	
	Diarrhoea	36.54	10.19	33.23-39.84	1.52
TLC (thousands/cm <sup>3</sup> )	Non-diarrhoea	8.15	3.81	5.59-10.71	0.03
	Diarrhoea	8.12	3.16	7.09-9.14	
TEC (million/cm <sup>3</sup> )	Non-diarrhoea	4.95	2.87	3.03-6.88	
	Diarrhoea	5.32	2.36	5.56-6.09	0.39
Lymphocytes (%)	Non-diarrhoea	69.45	8.91	63.47-75.44	
	Diarrhoea	71.72	10.05	68.46-74.97	0.72
Neutrophils (%)	Non-diarrhoea	12.09	8.43	6.43-17.76	
	Diarrhoea	19.79	11.52	16.06-23.53	2.45
Monocyte (%)	Non-diarrhoea	5.54	3.67	3.08-8.01	1.12
	Diarrhoea	4.23	2.47	3.43-5.03	
Basophil (%)	Non-diarrhoea	0.09	0.30	0.11-0.29	
	Diarrhoea	0.05	0.22	0.02-0.12	

 Table 3: Association of hematological parameters for non-diarrheic with diarrheic.

Variable	Infection status	Mean	SD	95% CI	P-value (Ttest)
Sodium (mEq/litre)	Non-diarrhoea	152.52	27.06	134.34-170.70	2.47
	Diarrhoea	128.59	18.87	122.47-134.71	
Potassium (mEq/litre)	Non-diarrhoea	5.74	0.94	5.10-6.37	
	Diarrhoea	6.23	1.44	5.77-6.70	1.35
Chloride (mEq/litre)	Non-diarrhoea	99.55	9.21	93.36-105.74	
	Diarrhoea	115.05	19.18	108.83-121.27	3.74

Table 4: Association of blood biochemistry parameters for non-diarrheic with diarrheic.

#### Comparison of hematological and serological parameters of Rota virus positive and negative calves

The hemoglobin values in both rotavirus positive and negative cases irrespective of diarrheic and non-diarrheic calves were lower than the average normal range. The mean value of hemoglobin was recorded as 9.75 and 9.59 g/dl as in rotavirus positive and negative cases, respectively (Table 5). However, the association was not found significant (P-value = 0.82).

The PCV Values in both rotavirus positive and negative cases were lower than the average normal range. Average mean value of PCV was recorded as 36.33% and 33.64% in rotavirus positive and negative cases, respectively (Table 5). The P-value of the significant test was 0.38 (statistically non-significant).

The TEC Values in all the diarrheic calves were lower than the normal range. The mean values of the TEC were 4.44 and 5.65 million per cm<sup>3</sup> in both rotavirus positive and negative cases, respectively (Table 5); the TLC values in all the diarrheic calves were lower than the normal range. The mean values of TLC were 7.67 and 8.35 thousands per cm<sup>3</sup> in rotavirus positive and negative cases, respectively.

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359

360

In the present study, the lymphocyte count was observed higher in diarrheic calves as compared with the normal values and average mean value of lymphocyte was estimated as 75.58% and 68.96% in rotavirus positive and negative cases, respectively. On the other hand, the neutrophil count was lower in diarrheic calves as compared with the normal values and the average mean value appeared as 15.41% and 19.48% in rotavirus positive and negative cases, respectively. The monocyte count in the diarrheic calves was slightly higher in comparison to that of normal values of healthy calves, whereas, the mean value of monocyte count was 4.11% and 4.72% in rotavirus positive and negative cases, respectively (Table 5).

Variable	Infection status	Mean	SD	95% CI	P-value (Ttest)
Hemoglobin(g/dl)	Negative	9.75	2.40	8.89-10.60	0.82
	Positive	9.59	2.44	8.33-10.85	
PCV (%)	Negative	36.33	9.91	32.81-39.84	0.38
	Positive	33.64	10.85	28.06-39.22	
TLC (thousands/cm <sup>3</sup> )	Negative	8.35	3.42	7.14-9.56	0.49
	Positive	7.67	3.01	6.12-9.22	
TEC (million/cm <sup>3</sup> )	Negative	5.65	2.76	4.67-6.63	0.09
	Positive	4.44	1.42	3.69-5.19	
Lymphocytes (%)	Negative	68.96	10.18	65.35-72.58	0.02
	Positive	75.58	7.34	71.81-79.36	
Neutrophils (%)	Negative	19.48	12.66	14.99-23.97	0.23
	Positive	15.41	7.69	11.46-19.35	
Monocyte (%)	Negative	4.72	2.93	3.68-5.76	0.46
	Positive	4.11	2.49	2.83-5.40	
Basophil (%)	Negative	0.09	0.29	0.01-0.19	0.20
	Positive	0	0	0	

**Table 5:** Association of hematological parameters with rota virus positive and negative calves diagnosed by rtRT-PCR.

Three common electrolytes (sodium, potassium and chloride) as a measure of homeostasis and water balance were studied and the mean values of the results are shown in Table 6. It can be noted that the sodium value in diarrheic calves was lower than that of normal values. Sodium was ranged between 132.28 and 134.66 mEq per litre in rotavirus positive and negative cases, respectively (Table 6). In the present study, it was recorded that the potassium values in diarrheic calves were slightly higher than that of normal values. The mean values of potassium in rotavirus positive and negative cases were 5.81 and 6.28 mEq per litre, respectively. Similarly, estimated value of chloride was higher in the diarrheic calves as compared to that of normal values. The mean values of chloride in rotavirus positive and negative cases were 109.42 and 115.93 mEq per litre, respectively. However, none of the significance test showed any significant association between different biochemical parameter and diarrheic or rota virus positive calves.

#### Comparison of hematological and serological parameters in diarrheic rota virus positive and negative calves

The results of hematological and serological parameters for diarrheal calves with rotavirus positive and negative cases represented in the table 7 & 8. The average lymphocyte value of rotavirus positive diarrheal calves were 74.69% which were higher than the value of rotavirus negative diarrheal calves that represented 69.65% (Table 7).

Variable	Infection status	Mean	SD	95% CI	P-value (Ttest)
Sodium (mEq/litre)	Negative	134.66	25.39	125.66-143.68	0.73
	Positive	132.28	17.80	123.12-141.43	
Potassium (mEq/litre)	Negative	6.28	1.40	5.78-6.77	0.25
	Positive	5.81	1.25	5.17-6.46	
Chloride (mEq/litre)	Negative	109.42	16.71	103.49-115.35	0.24
	Positive	115.92	21.61	104.81-127.04	

**Table 6:** Association of blood biochemistry parameters with

 rotavirus positive and negative calves diagnosed by rtRT-PCR.

Variable	Infection status	Mean	SD	95% CI	P-value (Ttest)
Hemoglobin (g/dl)	Negative	10.4	2.35	9.38-11.41	0.72
	Positive	9.86	2.27	8.65-11.07	
PCV (%)	Negative	38	9.80	33.76-42.24	1.06
	Positive	34.44	10.69	28.74-40.13	
TLC (thousands/cm <sup>3</sup> )	Negative	8.71	3.45	7.22-10.20	1.51
	Positive	7.26	2.57	5.89-8.63	
TEC (million/cm <sup>3</sup> )	Negative	5.84	2.76	4.64-7.03	1.87
	Positive	4.58	1.38	3.85-5.32	
Lymphocytes (%)	Negative	69.65	11.59	64.64-74.66	
	Positive	74.69	6.54	71.20-78.17	1.73
Neutrophils (%)	Negative	22.39	13.21	16.68-28.10	1.91
	Positive	16.06	7.42	12.11-20.02	
Monocyte (%)	Negative	4.22	2.49	3.14-5.29	
	Positive	4.25	2.52	2.91-5.59	0.04
Basophil (%)	Negative	0.09	0.29	0.04-0.21	1.45
	Positive	0	0	0	

**Table 7:** Association of hematological parameters for diarrhea with

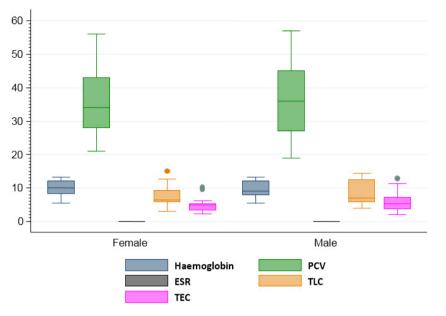
 rota virus positive and negative calves diagnosed by rtRT-PCR.

Variable	Infection status	Mean	SD	95% CI	P-value (Ttest)
Sodium (mEq/litre)	Negative	127.41	20.74	118.44-136.38	
	Positive	130.29	16.32	121.59-138.99	0.49
Potassium (mEq/litre)	Negative	6.51	1.51	5.86-7.16	1.49
	Positive	5.84	1.29	5.150-6.52	
Chloride (mEq/litre)	Negative	114.23	17.18	106.81-121.67	
	Positive	116.22	22.29	104.34-128.10	0.30

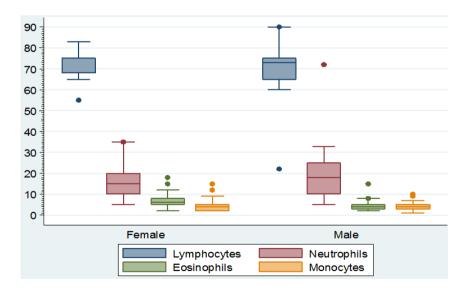
**Table 8:** Association of blood biochemistry parameters for diarrhoea

 with rota virus positive and negative calves diagnosed by rtRT-PCR.

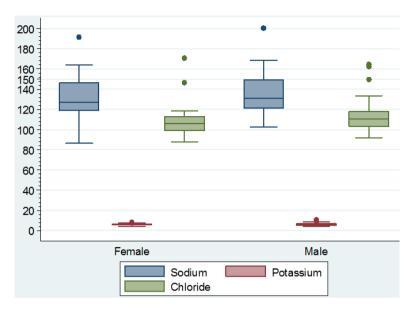
Visual differences in the maximum, minimum and median values of hemoglobin, PCV, ESR, TLC and TEC, lymphocytes, neutrophils, eosinophil's and monocytes according to sex of the study population can be noted in figure 1 & 2. Similarly, the maximum, minimum and median value of sodium, potassium and chloride according to sex of the study population showed visual differences in figure 3.



*Figure 1:* Boxplot showing the maximum, minimum and median value of haemoglobin, PCV, ESR, TLC and TEC according to sex of the study population.



*Figure 2:* Boxplot showing the maximum, minimum and median value of lymphocytes, neutrophils, eosinophil's and monocytes according to sex of the study population.



*Figure 3:* Boxplot showing the maximum, minimum and median value of sodium, potassium and chloride according to sex of the study population.

#### Discussion

In the present study, the hemoglobin value (Table 3, 5 &7) was lower in the study population compared to the reference value (Table 1) but it was higher in diarrheic than in non-diarrheic calves. The increase in hemoglobin in diarrheic calves might be due to haemoconcentration associated with dehydration. Our finding was in concordant with the findings of Schalm., *et al.* (1975) and [19, 23].

The elevation of PCV was observed in diarrheic calves compared to non-diarrheic. The increased PCV values in diarrheic calves indicate fluid loss from vascular compartment [13]. The elevation of PCV is an indicator of dehydration [24,25]. Thus, estimation of PCV is of utmost importance to monitor hydration status of animal and is a sensitive indicator for assessing the severity of dehydration. Increase in PCV in diarrheic calves was apparently due to hemo-concentration associated with dehydration and hypovolemia.

Elevated TEC was revealed in the diarrheic calves in the present study. The result is supported by the earlier findings by Schalm., *et al.* (1975) and [19] reported significant elevation of TEC in diarrheic calves. Haemoconcentration of blood due to diarrhea might be the underlying reason for this trend.

TLC value did not show any difference between diarrheic and non-diarrheic calves in the present study. Different studies revealed that the value of TLC increased in diarrheic calves and also reported significant leucocytosis which might have occurred due to normal reaction of body defense mechanism against infective cause of diarrhea and also due to haemoconcentration due to dehydration [19,26].

Differential leukocyte count (DLC) includes the estimation of relative percentage of neutrophils, eosinophil's, basophils, lymphocytes and monocytes in per ml of blood. Increase in DLC is suggestive of infectious agents of different origin. The present study revealed increase in lymphocyte count in rotavirus positive cases than rotavirus negative indicates a viral etiology of the diarrhea (Table 5 & 7).

The sodium is the most important cation in extracellular fluid as it is responsible for maintenance of osmotic pressure[10]. Together with chlorine (Cl) it collaborates in metabolism of water and regulation of acid-base balance in the cell [10]. The present study revealed significant reduction of Na values in diarrheic calves. These findings were in agreement with observations of [19,27,28]. Similarly, [29] also reported hypernatremia in diarrheic calves due to an excessive secretion of sodium along with water into intestinal lumen. These

findings are contrary to that of [30] who recorded hypernatraemia in diarrhoeic calves as compared to healthy calves. Most of the diarrhoea causing microorganisms disrupt the intestinal function and dehydrate the body either by increasing the chloride-secreting activity of the crypt cell or impairing the absorption of sodium by the villus cells or both [31].

Potassium is important for making electrical potential for transport of nerve impulses and for maintenance of muscle tonicity [10]. Potassium is also important for regulation of acid-base balance in the body [10]. Hypokalemia increase the membrane potential and cause hyper polarization block, that has influence on lower muscle tone and paralysis [32]. The present study revealed elevation of potassium levels in all the diarrheic calves. Hyperkalemia is caused by increased potassium retention by kidney and also due to cellular damage [33,34]. Hyperkalemia observed in the present study might be due to damage of enterocytes by microorganisms. Similar observations have been reported by other researchers [19,30,35]. In our study, serum chloride values of diarrheic calves showed elevation of chloride values in diarrheic calves.

## Conclusion

It can be concluded that diarrheal condition of calves influences the hematological and serological variables that could be considered during diagnosis. In the study, though, no statistically significant alterations were observed between the diarrheic and non-diarrheic calves, the analysis of blood and serum samples of the diarrheic calves revealed the elevation of Hb, PCV, TLC and Chloride compared to non-diarrheal calves. Similarly, Rota viral positive diarrheal cases showed elevation in lymphocyte count along with some other parameters. Finally, the results of the study can be used to partially support the diagnosis of the rotavirus infection in calves.

#### Acknowledgement

This work is supported by University Grants Commission Higher Education and Quality Enhancement Project (CP-3220).

#### **Authors Contribution**

SRB, TMR designed and conducted the research. SC, MM, MAH supervised the research. SD supported in manuscript preparation.

## **Conflict of interest**

Authors have no conflict of interest.

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